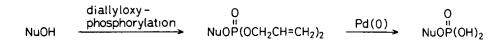
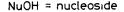
A GENERAL APPROACH TO NUCLEOSIDE 3'- AND 5'-MONOPHOSPHATES

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Summary : Diallyloxyphosphorylation of nucleoside hydroxyls followed by palladium(0)-catalyzed deallylation provides a new, general method for the preparation of the 3'- and 5'-monophosphates.

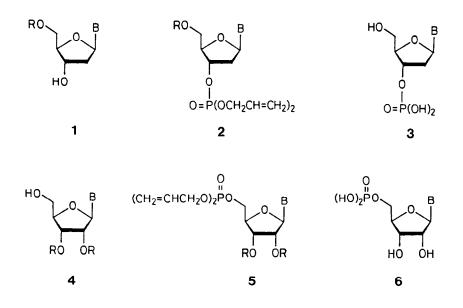
The 3'- and 5'-O-monophosphorylation of nucleosides is the significant problem in nucleotide synthesis because the monophosphates serve as the key synthetic intermediates of biologically important nucleoside di- and triphosphates.¹ However, most of the chemical and enzymatic methods developed up to now are applicable only to the 5'-O-phosphorylation.^{2,3} The appropriate way to phosphorylate the 3'-hydroxyls is still elusive.⁴ We disclose herein a general entry to the monophosphates via palladium(0)-mediated deprotection⁵ of the diallyl-blocked precursors.





The diallyloxyphosphorylation of the hydroxyls, the key step of the present method, is achievable in two different ways. One approach is the Grignard reagent-promoted coupling of nucleosides and diallyl phosphorochloridate⁶ (method A).⁷ For instance, when the 3'-O-free nucleoside 1 [B = Cy; R = t-C₄H₉(CH₃)₂Si (TBDMS)] was treated with one equiv of *t*-butylmagnesium chloride and then the diallyloxyphosphorylating agent at room temperature for 0.3 h, the protected nucleoside phosphate **2** (B = Cy; R = TBDMS) was obtained in 97% isolated yield. The other way to prepare the diallyl phosphates is base-assisted condensation of nucleosides and diallyl phosphorochloridite⁸ and oxidation of the resulting phosphite intermediates (method B). Thus, reaction of **1** [B = Th; R = C₆H₅(p-CH₃OC₆H₄O)₂C (DMTr)] and the phosphorochloridite in the presence of 2, 4, 6-collidine (-78 °C, 0.5 h) followed by oxidation by *t*-butyl hydroperoxide⁹ (0 °C, 20 min) gave the phosphorotriester **2** (B = Th; R = DMTr) in 89% yield. Similarly, the 5'-O-diallyloxyphosphorylation of **4** was accomplished according to either method. These methods are quite general and applicable to all kinds of nucleosides. Several examples are listed in Table I. Although the reaction using *N*-free cytidine or guanosine derivatives by method B did not give satisfactory yields of the diallyloxyphosphorylated products, the protection of the nucleoside bases improved the efficiency to a considerable extent. We recommend method A for the reaction of simple *N*-unprotected cytidines and guanosines.

Removal of the allyl and other protective groups allows the conversion of the phosphorotriesters, 2 and 5, to the desired nucleoside 3'-phosphate 3 and 5'-phosphate 6, respectively. The way of the deblocking was dependent on the



nucleoside		condensation		% yield of 3'-or 5'-
number (B)	R or R—R	method	time/h	phosphate ^a
1 (Ad)	DMTr	А	1	98
1 (Ad)	DMTr	B	4	87
1 (Cy)	TBDMS	А	0.3	97
1 (Cy)	TBDMS	В	2.3	64
1 (AOC-Cy ^b)	DMTr	В	0.5	92
1 (Gu)	TBDMS	\mathbf{A}^{c}	0.7	94
1 (Gu)	TBDMS	В	1.3	37
1 (AOC-Gu ^d)	DMTr	В	1	70 ^e
1 (Th)	DMTr	\mathbf{A}^{c}	1	99
1 (Th)	DMTr	В	0.5	89
4 (Ad)	(CH ₃) ₂ C	А	2	90
4 (Ad)	(CH ₃) ₂ C	В	1	93
4 (Cy)	(CH ₃) ₂ C	А	1	91
4 (Gu)	TBDMS	A ^c	1	93
4 (Ur)	(CH ₃) ₂ C	\mathbf{A}^{c}	4.5	86
4 (Ur)	(CH ₃) ₂ C	В	0.7	94

^a Isolated yield. ^b N^{4} -Allyloxycarbonylcytosinyl ^c Two equivalents of t-butylmagnesium chloride was employed. ^a N^{2} ıllyloxycarbonylguanyl. ^e Yield based on the consumed starting material (ca. 80%). mode of the protection, and the suitable reaction sequences and conditions were as follows: For the allyl—trityl derivatives, (1) CHCl₂COOH in CH₂Cl₂, 25 °C, 10 min, (2) 10 mol % Pd[P(C₆H₅)₃]₄, 60 mol % P(C₆H₅)₃, and n-C₄H₉NH₂—HCOOH (1:4, excess) in THF, 50 °C (method C);¹⁰ for the allyl—silyl derivatives, (1) 10 mol % Pd[P(C₆H₅)₃]₄, 60 mol % P(C₆H₅)₃]₄, 60 mol % P(C₆H₅)₃, and n-C₄H₉NH₂—HCOOH (1:4, excess) in THF, 50 °C, (2) CHCl₂COOH in CH₂Cl₂, 25 °C, 2—10 min (method E); for the allyl—isopropylidene derivatives, (1) 10 mol % Pd[P(C₆H₅)₃]₄, 60 mol % P(C₆H₅)₃, and n-C₄H₉NH₂—HCOOH (1:4, excess) in THF, 50 °C, (2) 2 N aq HCl, 25 °C, 12 h (method F). Table II summarizes some examples of the deprotection. The overall process left the whole nucleotide functionalities completely intact.

The present approach provides the first general way capable of preparing all kinds of 2'-deoxyribonucleoside 3'phosphates, and is advantageous over the existing methods in its applicability to a wide range of nucleosides.

nucleotide		deprotection		0/ 111 (0 0/
number (B)	R or R-R	method	Pd treatment/h	% yield of 3 or 6^{α}
2 (Ad)	DMTr	С	3.7	91
2 (Cy)	TBDMS	D	3	91
2 (AOC-Cy ^b)	DMTr	E	4.3	90
2 (Gu)	TBDMS	D	4.3	91
2 (AOC-Gu ^c)	DMTr	Е	4	83
2 (Th)	DMTr	С	4.3	90
5 (Ad)	$(CH_3)_2C$	F	5	89
5 (Cy)	(CH ₃) ₂ C	F	5.7	92
5 (Gu)	TBDMS	D	5.3	100
5 (Ur)	(CH ₃) ₂ C	F	5.7	91

Table II. Deprotection of the Nucleoside Phosphorotriesters, 2 and 5

^a HPLC yield of the overall deprotection. ^b N⁴-Allyloxycarbonylcytosinyl ^c N²-Allyloxycarbonylguanyl

Diallyloxyphosphorylation of Nucleosides by Method A. To a solution of 1 or 4 (0.5 mmol) in THF (10 mL) was added a THF solution of *t*-butylmagnesium chloride (1 equiv for B = Ad, Cy; 2 equiv for B = Gu, Th, Ur) at ambient temperature. After 10 min, diallyl phosphorochloridate (0.5 mmol) was added and the resulting mixture was stirred for the stated period. Usual extractive workup followed by chromatography on silica gel (CH₃OH/CHCl₃ = 1/50 to 1/20) gave the phosphorotriester 2 or 5, respectively.

Diallyloxyphosphorylation of Nucleosides by Method B. A mixture of the nucleoside, 1 or 4 (0.2 mmol), diallyl phosphorochloridite (1.2 mmol), and 2, 4, 6-collidine (1.0 mmol) in THF (1 mL) was stirred at -78 °C for the stated period. The mixture was quenched by addition of methanol (0.1 mL) at -78 °C and warmed to 0 °C. To this was added a 1.9 *M* toluene solution of *t*-butyl hydroperoxide (2.0 mmol) and stirring was continued for 20 min. Extractive workup and subsequent chromatography as described above afforded 2 or 5, respectively.

Typical Example of Deprotection. A mixture of 2 (B = Cy, R = TBDMS) (89 mg, 0.18 mmol), Pd[P(C₆H₅)₃]₄ (20 mg, 0.018 mmol), P(C₆H₅)₃ (27 mg, 0.11 mmol), butylamine (70 μ L, 0.71 mmol), and formic acid (106 μ L, 2.8 mmol) in THF (2

mL) was kept at 50 °C with stirring for 3 h. Concentration gave an oil and this was then treated with a 1 *M* THF solution of TBAF (10 mL, 10 mmol) at ambient temperature for 1 h. Removal of the solvent *in vacuo* gave a viscous residue, which was subjected to HPLC analysis ODS-5 μ m, 0.1 *M* KH₂PO₄ + 0.005 *M* (*n*-C₄H₉)₄NBr aq buffer/CH₃OH = 95/ 5, 260 nm, 1 mL/min, 40 °C, indicating the formation of 3 (B = Cy) in 91% yield. DEAE cellulose (DE52) column chromatography[HCO₃⁻ form, 3.8 x 12 cm, H₂O to 0.15–1 *M* triethylammonium hydrogencarbonate (pH 7.4)] of the crude material afforded triethylammonium salt of the nucleotide (1436 OD₂₇₀ units, 89% yield) in a pure form.

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(Received in Japan 5 February 1987)